





Interim Advice for the Diagnosis and Management of PVL-associated Staphylococcus aureus infections (PVL-S. aureus)

Scottish Recommendations

May 2014



Document Amendment Log

| Version No. | Date | Page No. | Amendment Summary |
|----------------|------|----------|-------------------|
| | | | |
| | | | |
| | | | |
| | | | |
| | | | |
| | | | |

The Health Protection Network (HPN) is a network of existing professional organisations and networks in the health protection community across Scotland. It aims to promote, sustain, and coordinate good practice. The HPN supports a systematic approach to development, appraisal and adaptation of guidelines, seeking excellence in health protection practice.

Health Protection Network site: <u>http://www.hps.scot.nhs.uk/about/HPN.aspx</u>.

Supported by Health Protection Scotland

Health Protection Scotland (HPS) is a non-profit, public sector organisation which is part of the Scottish National Health Service. It is dedicated to the protection of the public's health.

Health Protection Scotland is a division of NHS National Services Scotland.

Reference this document as:

Health Protection Network. Interim Advice for the Diagnosis and Management of PVLassociated Staphylococcus aureus infections (PVL-S. aureus). Health Protection Network Scottish Guidance 10. Health Protection Scotland, Glasgow, 2014.

Published by Health Protection Scotland Meridian Court, 5 Cadogan Street, Glasgow, G2 6QE.

First published May 2014.

© Health Protection Network 2014.

The Health Protection Network has made every effort to trace holders of copyright in original material and to seek permission for its use in this document and in any of the associated quick reference guide. Should copyrighted material have been inadvertently used without appropriate attribution or permission, the copyright holders are asked to contact the Health Protection Network so that suitable acknowledgement can be made at the first opportunity.

Health Protection Network consents to the photocopying of this document for the purpose of implementation in NHSScotland.

All other proposals for reproduction of large extracts should be addressed to:

Health Protection Network Health Protection Scotland Meridian Court 5 Cadogan Street Glasgow G2 6QE

Tel: +44 (0) 141 300 1100

Email: <u>nss.hpsenquiries@nhs.net</u>

Professionals involved in the implementation of recommendations proposed in this document are expected to take them fully into account when exercising their professional judgment. The document does not, however, override the individual responsibility of professionals to make decisions appropriate to the circumstances of the individual cases, in consultation with partner agencies and stakeholders. Professionals are also reminded that it is their responsibility to interpret and implement these recommendations in their local context, in light of their duties to avoid unlawful discrimination and to have regard to promoting equality of opportunity. Nothing in this document should be interpreted in a way which would be inconsistent with compliance with those duties.

Designed and typeset by:

Graphics Team, Health Protection Scotland

Table of Contents

| Abb | revio | ations | | 1 |
|-----|-------------|--------------------|--|----|
| 1. | Abo | ut this | Document | 2 |
| 2. | Bac | kgrour | nd and Epidemiology | 4 |
| 3. | Clin | ical fe | atures and Diagnosis of PVL-S. aureus Infections | 6 |
| | 3.1 | Diagno Pneum | osis of Patients with Community-Acquired (CA) Necrotising nonia | 7 |
| | 3.2 | Risk Fa | ictors for PVL-S. aureus Infections Transmission | 8 |
| 4. | Micı | robiolo | ogical Samples and PVL Testing | 9 |
| | 4.1 | PVL Te | sting in SMRSARL | 9 |
| 5. | Mar | nagem | ent of Suspected and Confirmed PVL-S. aureus Cases | 10 |
| | 5.1 | Antibio Infecti | otic Treatment of Suspected and Confirmed PVL-S. aureus ons in Adults | 10 |
| | 5.2 | Specie Suspec | al Consideration Around the Management of Confirmed and cted PVL-S. <i>aureus</i> Infections in Children | 14 |
| 6. | Scre | ening | and Decolonisation of Patients and their Close Contacts | 16 |
| | 6.1 | Screer | ning of Patients and their Close Contacts | 16 |
| | 6.2 | Decol | onisation of Patients and Contacts | 16 |
| 7. | Clus | ters ar | nd Outbreaks of PVL-S. aureus Infections | 18 |
| 8. | Infe | ction P | revention and Control (IPC) Measures | 20 |
| | 8.1 | Infecti | on Prevention and Control Measures for Hospitalised Patients | 20 |
| | | 8.1.1 | Infection Prevention and Control (IPC) Measures in Relation to Patients with Necrotising Pneumonia | 20 |
| | | 8.1.2 | IPC Measures in Relation to Hospital-acquired Infections | 21 |
| | | 8.1.3 | Occupational Health | 21 |
| | 8.2 | Infecti | on Prevention and Control Measures for Household Settings | 22 |
| Арр | endi (Ad | x A: Q apted | uick Reference Guide for PVL-S. <i>aureus</i> for Primary Care from HPA Chart 18/5/09) | 29 |
| Арр | endi | x B: Pc | atient information – Decolonisation General Information | 31 |
| Арр | endi | x C: Po | atient Information PVL-Staphylococcus aureus | 32 |
| Арр | endi | x D: G | uideline Development Group (GDG) Membership | 34 |
| Арр | endi | X E: HP | N Guideline Feedback form | 36 |

Abbreviations

| BNF | British National Formulary |
|---|--|
| BNFc | British National Formulary for children |
| CA | Community Acquired |
| CA-MRSA | Community-associated meticillin-resistant Staphylococcus aureus |
| CRP | C- Reactive Protein |
| ESR | Erythrocyte Sedimentation Rate |
| GDG | Guideline Development Group |
| GP | General Practitioner |
| HA-MRSA | Hospital-associated meticillin-resistant Staphylococcus aureus |
| HCW | Healthcare worker |
| HPA | Health Protection Agency |
| HPS | Health Protection Scotland |
| HPT | Health Protection Team |
| ICT | Infection Control Team |
| IPC | Infection Prevention Control |
| IMT | Incident Management Team |
| IV | Intravenous |
| | |
| IVIG | Intravenous immunoglobulin |
| IVIG MRSA | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus |
| IVIG MRSA MSSA | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus |
| IVIG MRSA MSSA PAG | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus Problem Assessment Groups |
| IVIG MRSA MSSA PAG PHE | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus Problem Assessment Groups Public Health England |
| IVIG MRSA MSSA PAG PHE PVL | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus Problem Assessment Groups Public Health England Panton-Valentine Leukocidin |
| IVIG MRSA MSSA PAG PHE PVL SA | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus Problem Assessment Groups Public Health England Panton-Valentine Leukocidin Staphylococcus aureus |
| IVIG MRSA MSSA PAG PHE PVL SA SICP | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus Problem Assessment Groups Public Health England Panton-Valentine Leukocidin Staphylococcus aureus Standard Infection Control Precautions |
| IVIG MRSA MSSA PAG PHE PVL SA SICP SSTI | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus Problem Assessment Groups Public Health England Panton-Valentine Leukocidin Staphylococcus aureus Standard Infection Control Precautions Skin and soft tissue infection |
| IVIG MRSA MSSA PAG PHE PVL SA SICP SSTI RCPI | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus Problem Assessment Groups Public Health England Panton-Valentine Leukocidin Staphylococcus aureus Standard Infection Control Precautions Skin and soft tissue infection Royal College of Physicians of Ireland |
| IVIG MRSA MSSA PAG PHE PVL SA SICP SSTI RCPI TSS | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus Problem Assessment Groups Public Health England Panton-Valentine Leukocidin Staphylococcus aureus Standard Infection Control Precautions Skin and soft tissue infection Royal College of Physicians of Ireland Toxic shock syndrome |
| IVIG MRSA MSSA PAG PHE PVL SA SICP SSTI RCPI TSS TBP | Intravenous immunoglobulin Meticillin-resistant Staphylococcus aureus Meticillin-sensitive Staphylococcus aureus Problem Assessment Groups Public Health England Panton-Valentine Leukocidin Staphylococcus aureus Standard Infection Control Precautions Skin and soft tissue infection Royal College of Physicians of Ireland Toxic shock syndrome Transmission Based Precautions |

1. About this Document

The recommendations in this document have been produced by the PVL Working Group – Guideline Development Group (GDG) (see <u>section Appendix D: Guideline</u> <u>Development Group (GDG) Membership on page 34</u>) convened in 2012, at the request of the Scottish Health Protection Network (HPN) Steering Group.

- This document aims to provide advice based on best available evidence where it exists (otherwise expert consensus opinion), on the recognition, investigation and management of PVL Staphylococcus aureus (PVL-S. aureus) cases in Scotland. It aims to assist all healthcare professionals who are involved in the management of these patients, from Primary Care to Public Health.
- This document is based on the HPA 'Guidance on the Diagnosis and Management of PVL-associated Staphylococcus aureus infections' (HPA, 2008) and some sections have been reproduced with permission from Public Health England (PHE).
- In 2011, the HPN commissioned the appraisal of the HPA 2008 document to independent external reviewers. The outcome of that appraisal (using the AGREE Instrument, 2009), as well as a subsequent review of the literature published before October 2012, further informed the GDG, who in the last year extensively consulted key partners and experts in the field.
- Recommendations given in this guidance are the result of careful review and consideration of the evidence available, existing guidance, expert opinion and principles of best practice.
- Public Health England (PHE) is currently, at the time this has been published, reviewing their 2008 guidance. As developments in the field have occurred, it has become apparent that advice is required not only for the management of PVL-S. aureus but for that of a wider suite of virulence determinants for Staphylococcus aureus and their resultant clinical syndromes. The UK GDG, therefore, has an overall aim of producing UK-wide evidence-based clinical guidance on the prevention and management of toxigenic Staphylococcus aureus (TSA) infections, excluding enterotoxigenic disease.
- The Health Protection Network (HPN) is represented at the UK guideline development group. Therefore, this document is an interim document that will provide advice for Scotland, while the UK group works on a more systematic approach to review the updated evidence. The HPN will review the recommendations proposed by the UK group and decide on the most appropriate advice for Scotland.

Professional Judgement and Compliance to this Document

Professionals involved in the investigation and management of PVL-S. aureus infections in Scotland are expected to take this guidance document fully into account when exercising their professional judgment. The document does not, however, override the individual responsibility of professionals to make decisions appropriate to the circumstances of the individual incidents and cases, in consultation with partner agencies and stakeholders. Implementation of this guidance is the responsibility of the health protection community across Scotland. Professionals are reminded that it is their responsibility to implement the recommendations provided, in their local context, in light of their duties to avoid unlawful discrimination and to have regard to promoting equality of opportunity. Nothing in this guidance should therefore be interpreted in a way which would be inconsistent with compliance with those duties.

Comments on the Published Guidance

Comments on this document should be sent to the HPN Steering Group via its national coordinator or administration, submitting the form available at the http://www.hps.scot.nhs.uk/about/guidancedevelopment.aspx, to the following email address http://www.hps.scot.nhs.uk/about/guidancedevelopment.aspx.

Sometimes a comment after publication may highlight a potential error in clinical guidance. This might be in either the interpretation or the presentation of the evidence considered by the GDG. In these cases the Chair of the Health Protection Network and the advisors they approach will consider whether the potential error:

- may result in harm to patients/the population;
- undermines the conclusions on which the recommendations were based;
- indicates serious problems with our quality-assurance procedures.

If one of these criteria is met, the comment will be referred to the HPN Guidance Executive, which decides what action to take. If the Guidance Executive does not accept that an error has been made, the individual or organisation that made the comment will be notified. If the Guidance Executive accepts that an error has been made, a note will be put on our website, and the versions of the document on the website will be amended. Depending on the nature and significance of the error and the time since publication, registered stakeholders may also be notified in writing.

Comments or new evidence which do not relate to an error will be collated and considered at the time when review of the document is due.

Review and Update

This document should be reviewed in 3 years or when the UK national guidance becomes available, whichever is sooner. It is envisaged that the HPN will oversee this process.

2. Background and Epidemiology

Panton-Valentine Leuocidin (PVL) is a cytotoxin that destroys white blood cells and is produced by some strains of *Staphylococcus aureus* (Etienne and Dumitrescu, 2009).

PVL takes its name from two authors who first published a paper in 1932 (Panton, 1932) and is encoded by two genes luk-S-PV and luk-F-PV. PVL is transferred by bacteriophages in some heterogeneous strains of Meticillin-Resistant *Staphylococcus aureus* (MRSA) and Meticillin-Sensitive *Staphylococcus aureus* (MSSA). It should be noted that terminology is changing and PVL has also been referred to under the name Toxigenic *Staphylococcus aureus* (TSA).

There is currently a lack of robust evidence on prevalence and incidence of PVL *S. aureus* infection (Coia *et al*, 2006). However, over the last decade, there have been multiple case reports from the UK, Europe, USA and Asia of patients with various forms of the disease, indicating that PVL-*S. aureus* is on the increase (Ellington MJ *et al*, 2009; Koch R, 2010; Nhan TX *et al*, 2012).

The increase in morbidity and mortality associated with PVL-MRSA has caused public health concern worldwide. To date most PVL-*S*. *aureus* strains in the UK have been MSSA, but a major problem has emerged with Community Acquired MRSA (CAMRSA) in North America, most of which produce PVL. One strain in particular, the USA300 clone, is now spreading in hospitals in the USA (Patel, 2008).

PVL-S. aureus infections, although currently uncommon in the UK, seem to have increased in the last few years. It is not clear, however, whether the increasing numbers observed reflect improved case ascertainment of PVL-related syndromes and/or an increasing real prevalence of PVL-S. aureus.

From a UK perspective, fatalities due to PVL-S. *aureus*, although rare, and seen in both community and healthcare settings, have attracted high-profile media attention and prompted concern regarding the transmissibility and virulence sometimes associated with these organisms (Figure 1). PVL-S. *aureus* has emerging epidemiology so this document is based on literature up to and including October 2012. Further details on epidemiology in Scotland can be accessed via the Scottish MRSA Reference Laboratory and/or Health Protection Scotland (HPS). FIGURE 1: Clinical iceberg of Panton-Valentine leucocidin-associated infection - with permission from Shallcross LJ *et al* (2013)



3. Clinical features and Diagnosis of PVL-S. aureus Infections

PVL-S. aureus predominantly cause skin and soft tissue infections (SSTI) (Rajendran PM, 2007), but can also cause invasive infections (Morgan, 2007), the most serious of these is a necrotising haemorrhagic pneumonia with a high mortality (Gillet Y, 2002). This often follows a 'flu-like' illness, and may affect otherwise healthy young people in the community as well as those of any age with underlying conditions that may predispose them to staphylococcal infection. The clinical presentation of PVL-S. aureus infection is summarised in table 1.

| Skin and soft tissue infections (SSTI) | Invasive infections |
|---|---|
| These are often recurrent and include: • Boils (furunculosis), carbuncles, | Necrotising pneumonia (McGrath B, 2008) |
| folliculitis, cellulitis, purulent eyelid | Necrotising fasciitis |
| infection | Osteomyelitis, septic arthritis, and |
| Tissue necrosis | pyomyositis |
| Abscesses | Purpura fulminans |
| Fogo A, Kemp N, Morris-Jones R (2011) | • Bacteraemia |

TABLE 1: Clinical presentation of PVL-S. aureus infections.

In addition to the clinical presentation (as above), PVL-S. aureus infections should be suspected:

- when there are clusters of skin or soft tissue infections within a household or social group;
- when community-acquired necrotising/haemorrhagic pneumonia presents in young, previously fit people.

PVL infections in children are associated (in particular) with:

- enhanced inflammatory response (higher ESR and C-reactive protein (CRP));
- local disease (myositis/pyomyositis) and multiple site/recurrent furunculosis;
- severe sepsis that may present with purpura fulminans and/or deep vein thrombosis;
- pneumonia that may present with preceding 'flu-like' illness; haemoptysis, leuco/neutropenia and multilobular infiltrates;
- acute haematogenous osteomyelitis or osteoarticular infections (Bocchini EC, Hulten KG, et al, 2006).

3.1 Diagnosis of Patients with Community-Acquired (CA) Necrotising Pneumonia

Early clinical diagnosis is difficult but essential for improving survival. A household history of spreading or recurrent PVL-S. *aureus* skin sepsis may also be present (Morgan, 2007). **Table 4** provides a summary of the clinical parameters that may help identify community acquired (CA) necrotising pneumonia.

TABLE 2: Clinical parameters for the diagnosis of community acquired (CA) necrotising pneumonia.

| Clinical signs | Radiography | Clinical and Diagnostic Parameters |
|--|---|---|
| Pneumonia and sepsis in a previously fit young patient following a 'flu-like' illness warrant prompt referral to hospital if the following features are present: Airway bleeding/ haemoptysis; Hypotension; Tachycardia >140 beats/min, diarrhoea and vomiting (may be due to associated toxic shock). | Multilobular infiltrates on chest X-ray, usually accompanied by effusions and later cavitation. | Gram film of sputum reveals numerous Gram-positive cocci in grapelike clusters; Marked leucopenia (may be within normal limits early in illness as destruction by toxins is just beginning); Very high CRP level (>200g/L: unusual in viral infections); Negative Pneumococcal and Legionella antigen tests; Significantly raised serum creatine kinase (suggests myositis); Usual investigation blood cultures, viral testing, skin swabs etc to be taken. |

3.2 Risk Factors for PVL-S. aureus Infections Transmission

The risk factors for PVL-S. *aureus* transmission between individuals include: people in close contact (direct or indirect contact - via fomites).

The following settings may have higher risk for transmission from an individual colonised or infected with these staphylococci:

- household and sexual contacts;
- contacts in social/sports settings e.g. wrestling, American football, rugby, judo;
- closed community settings: e.g. military camps, gyms, prisons;
- carehomes and healthcare settings (hospital wards).

4. Microbiological Samples and PVL Testing

Clinical staff in primary and acute care settings should be alert to the importance of:

- further investigating suspected necrotising pneumonia (co-infection with a respiratory virus, including influenza);
- taking samples for culture and PVL toxin testing, when incising and draining multiple and recurring abscesses – as well as other samples as appropriate (e.g. blood cultures, swabs of pus, sputum, nasopharyngeal aspirate, etc);
- giving sufficient clinical details to enable laboratory staff to select any staphylococci for further testing; and
- discussing any clinical concerns with local Microbiologist or Infectious Disease Doctor.

4.1 PVL Testing in SMRSARL

In case of queries, the local Microbiology department can ask for advice from the Scottish MRSA Reference laboratory (SMRSARL) (contact details at <u>http://www.smrsarl.scot.nhs.uk/</u>):

- MSSA or MRSA isolated from PVL suspected cases should be referred to SMRSARL for toxin gene profiling, which includes PVL testing.
- All staphylococci from blood cultures are tested for PVL. This PCR-based assay is performed daily and completed within a working day. If results are needed urgently, these will be telephoned to the submitting laboratory if a request is made. Even if PVL testing is performed locally, isolates must be sent to SMRSARL for further toxin testing and typing, as this is currently the basis of national surveillance and provides early warning of changes in the national situation.

MRSAs with a typical susceptibility pattern for hospital type MRSAs and likely to have been acquired in a healthcare setting should not be referred unless clinical history suggests a PVL-S. aureus infection, e.g. necrotising pneumonia, recurrent boils etc. This information must be included on the referral forms. Ciprofloxacinresistant MRSA should not be referred for PVL testing unless they are associated with typical PVL-S. aureus related disease.

5. Management of Suspected and Confirmed PVL-S. aureus Cases

5.1 Antibiotic Treatment of Suspected and Confirmed PVL-S. aureus Infections in Adults (see <u>section 5.2 on</u> <u>page 14</u> for the management of children)

The following pattern should be adopted to treat suspected and confirmed PVL *S. aureus* infections in adults:

- Consult with the local Microbiologist;
- Consult any local antibiotic prescribing policies;
- Modify antibiotic choice, once sensitivity results are available.
- Inform local Infection Prevention and Control Team if the patient requires admission to hospital to ensure appropriate Transmission Based Precautions (TBPs) are put in place. (see <u>section 8. Infection Prevention and Control (IPC)</u> <u>Measures on page 20. Table 3</u> indicates generic advice in relation to the antibiotic treatment of suspected and confirmed infected adults. This advice should be considered in conjunction with risk assessment of each individual case and appropriate clinical judgement.

It should be noted that there are few robust clinical data on which to provide clear antimicrobial recommendations. The advice in <u>Table 3</u> reflects current experience, case reports and in vitro experimental data.

Current thinking is that flucloxacillin should be avoided in treatment of severe infections (even in combination with other antimicrobials or as high doses) as sub-inhibitory concentrations may enhance toxin production. In this situation, antimicrobials such as clindamycin or linezolid have the advantage of suppressing toxin production.

TABLE 3: Antibiotic Treatment of Suspected and Confirmed PVL-S. aureus Infections in Adults (see **section Table 4 on page 15** for children).

| Indications | Antimicrobial choice | Comments |
|--|---|--|
| Non-suppurative minor skin and soft tissue. Including furunculosis, folliculitis, small abscesses/boils without cellulitis. PVL - MRSA not suspected. | • Flucloxacillin orally if required | Does not need systemic antibiotic treatment unless the patient is immunocompromised, or deteriorating clinically. Incision and drainage is the optimal management for abscesses. Lesions should be covered. Advise good personal hygiene, in particular hand washing. Avoid sharing towels, cloths, personal care items. Patients should be advised to return to GP if the lesions do not resolve or there is clinical deterioration. |
| Moderate infections including cellulitis and larger abscesses (especially those > 5cm). PVL - MRSA not suspected. | Flucloxacillin OR Clindamycin OR Doxycycline Cotrimoxazole may also be used for penicillin allergic patients | Incision and drainage is the optimal management for abscesses. Note C. difficile risk with use of clindamycin. Review with laboratory antibiotic results when available. |
| When PVL-MRSA is suspected but not confirmed and hospital admission is not warranted. Liu C (2011) De Angelis G (2011) (*) Nathwani D (2008) | Doxycycline (not for children <12 y) OR Rifampicin PLUS Sodium Fusidate OR rifampicin PLUS trimethoprim OR Clindamycin | Resistance to Rifampicin and Fusidic acid may develop during treatment and these should not be used as single agents. Review with laboratory antibiotic results when available. Linezolid maybe available on advice from local Microbiologists or Infectious Disease Doctors. |

| Indications | Antimicrobial choice | Comments |
|--|--|---|
| PVL- MRSA confirmed | Treatment guided by antimicrobial susceptibility tests and local policy | Provide information to the patient – Patient Information leaflet (<u>section Appendix B</u> <u>on page 31</u> and <u>section</u> <u>Appendix C on page 32</u>). |
| In severe infections with features of toxic shock, necrotising fasciitis or purpura fulminans. Seek specialist advice from microbiology, infectious diseases, medical and surgical specialties as appropriate. | Refer to hospital There may be a theoretical case for using two agents such as these agents suppress toxin production: • Linezolid combined WITH Clindamycin (high dose). | Treatment should be continued for 10-14 days until the patient has improved and is clinically stable. Seek specialist advice from Microbiology or Infectious Diseases Team. Consult BNF for details on use of these antibiotics. Early surgical debridement of infected tissue where appropriate. Evidence from <i>in-vitro</i> synergy and the ability of linezolid and clindamycin to suppress PVL and alpha toxin production. Consider use of IV immunoglobulin using local protocols. Although bactericidal, there are concerns that at concentrations just above the minimum inhibitory concentration (likely with poor penetration into necrotic tissue) flucloxacillin may increase PVL production as it does <i>in-vitro</i> (Stevens et al, 2007). Intravenous flucloxacillin is not recommended, even in combinations with agents such as rifampicin or clindamycin. |

| Indications | Antimicrobial choice | Comments |
|--|---|--|
| In suspected community-acquired (CA) PVL-related pneumonia. | Start empiric antibiotics covering MRSA – linezolid and high dose of clindamycin. | Continue empiric antibiotic therapy for 48-72 hours or until culture results are available when targeted therapy can be consolidated: |
| Note: standard empiric antimicrobial cover for non staphylococcal pathogens may be required until microbiology results are available. | AND if deteriorating or features of severe disease (e.g. septic shock) add IVIG 2g/ kg + rifampicin 600 mg bd. | Seek advice from local Microbiologists or Infectious Disease Doctors. There is evidence that the use of rifampicin and linezolid combined may reduce blood concentration of linezolid. If no clinical improvements and increasing failure to ventilate: Exclude complications (e.g. abscess, empyema) and no infections – consider second dose of IVIG. Re-evaluate for infection with antibiotic-resistant pathogen not covered by initial antimicrobial regimen. |
| In case of deep- seated infections (e.g. Osteomyelitis/ disciitis) | Seek specialist advice | Seek specialist advice |

*<u>http://jac.oxfordjournals.org/content/61/5/976.full</u>

Table 3 should be followed in line with the following advice:

- Please consult BNF for adults (<u>http://www.medicinescomplete.com/mc/bnf/</u> <u>current/PHP3256-antibacterial-drugs.htm</u>) for advice on dose and dosage modifications if required with weight, renal and hepatic function, interactions and contraindications.
- Consult BNF for details on adverse and toxic events and monitoring requirements.
- Standard Infection Control Precautions must continue to be applied and additional TBPs considered and implemented as set out in the National Infection Prevention and Control Manual: <u>http://www.hps.scot.nhs.uk/haiic/</u> ic/nationalinfectionpreventionandcontrolmanual.aspx.

5.2 Special Consideration Around the Management of Confirmed and Suspected PVL-S. *aureus* Infections in Children

 Please consult BNF for Children for contraindications and precautions in the use of these antibiotics: <u>http://www.medicinescomplete.com/mc/bnfc/ current/PHP12608-infections.htm</u>.

Skin and soft tissue infections in children:

• These should be suspected if there are recurrent boils or abscesses or a close contact who has skin lesions.

Initial approach to therapy of deep-seated infections in children:

- Administer antibiotics according to local guidelines for empirical management of infections, but add clindamycin if PVL-MSSA is suspected. Linezolid can be used after discussion with Microbiologist or Infectious Disease Doctor;
- Close monitoring of clinical condition is essential, as some patients will deteriorate even after several days receiving appropriate antibiotic therapy;
- Ensure appropriate consultation, for instance with Medical Microbiologist, paediatric Infectious Disease Doctor, Orthopaedic Surgeon or Haematologist, as required. If patient admitted to hospital, inform Infection Prevention and Control Team so that appropriate measures are put in place.
- For the management of deep-seated infections in children, follow Table 4.

| Type of Illness | Indicators and Management |
|---------------------------------|---|
| Abscesses | Localised abscesses (e.g. retropharyneal or in lymph nodes) may be associated with local venous thrombosis, very high CRP and patient or close family contact has current, or a history of, recurrent boils/abscesses or skin infections. |
| Bone and joint infections | Suspect if patient or close family contact has current, or a history of, recurrent boils/abscesses or skin infections or there is severe sepsis, multiple sites of infection/abscesses, extensive local lesions, myositis/pyomyositis, local venous thrombosis, very high CRP and a need for repeated surgical intervention. Aggressive approach to drainage of foci of infection. Once infection with PVL-S. aureus is confirmed, use intravenous clindamycin (if susceptible) plus rifampicin, and consider use of linezolid or an alternative combination advised by a specialist in paediatric infectious disease. Repeated surgical intervention for drainage may be required, and the duration of antibiotic treatment may need to be very prolonged. Use of linezolid may be particularly useful in bone and joint infections. Linezolid should be used for a maximum of four-weeks due to the risk of development of peripheral neuropathy. For all antibiotics use the maximum dosages listed in the British National Formulary for children (BNFc) - http://www.medicines.complete.com/mc/bnfc/current/PHP12608-infections.htm. |
| Severe sepsis | Suspect if patient or close family contact has current, or a history of, recurrent boils/abscesses or skin infections, and there are bone or joint infection, necrotising pneumonia, deep venous thrombosis, purpura fulminans. consider transfer to paediatric ICU. consider 1-2g/kg IVIG and repeat lower dose after 24-48 hours if needed (see below) in addition to antibiotic treatment. |
| Pneumonia | Suspect if there is preceding 'flu-like' illness, haemoptysis, multilobular infiltrates, bone or joint infection, leucopenia/ neutropenia or patient or close family contact has current, or a history of, recurrent boils/abscesses or skin infections. Consider transfer to paediatric ICU. Consider 1-2g/kg IVIG and repeat lower dose after 24-48 hours if needed (see below) in addition to antibiotic treatment.* |

TABLE 4: Management of deep-seated infections in Children

* Use of IVIG: an initial dose of 1-2g/kg IVIG may be used in children, with some experts preferring a lower dose to reduce the risk of hyperviscosity occurring. This lower dose may be repeated after 24-48 hours if there is no clinical improvement.

6. Screening and Decolonisation of Patients and their Close Contacts

6.1 Screening of Patients and their Close Contacts

There is no evidence to support routine screening of contacts of patients with PVL-S. *aureus* infections (Shallcross LJ 2011). Patients and their household/sexual contacts should have high awareness of the potential for spread of PVL-S. *aureus*-related infections within their contact group:

- If close contacts suffer from infections typical or suggestive of PVL they should be advised to consult their GP for treatment, especially if they work or reside in high risk areas (see <u>section 3.2 on page 8</u>.), i.e. carehomes, closed communities (barracks, prisons) or healthcare settings.
- The decision to start contact screening in the case of a PVL-S. aureus outbreak within a healthcare setting will be risk assessed by the local Infection Prevention and Control Team.
- If the decision made concludes that screening is required, local MRSA screening procedures can be utilised in terms of which carriage sites to sample and the type of swabs to take.

6.2 Decolonisation of Patients and Contacts

The aims of a (topical) decolonisation programme for any staphylococcal infection include:

- reducing bacterial load for instance, pre-operatively;
- managing household outbreaks where recurrent infections in different household members has occurred;
- reducing the risk of infection in close contacts (household or sexual) of a case of necrotising pneumonia (start without delay); as well as
- interrupting transmission during clusters or outbreaks in 'closed' communities.
- reducing risk of onward transmission (e.g. healthcare workers (HCW), those in closed communities).

However, no data have been identified to date to support clinical or cost effectiveness of community-based screening and decolonisation for PVL strains (Shallcross LJ, 2013, 2011, Reilly, 2009).

It is important to observe that while decolonisation can be achieved temporarily, re-colonisation can occur relatively quickly. There is consistent evidence to demonstrate that two weeks after decolonisation, MRSA is undetectable in about 90% of those undertaking topical decolonisation regimens; at six months though, this drops to 30% (Ammerlaan HS, 2009).

With regards to resistance to topical decolonisation programmes, there are concerns about development of resistance to both chlorhexidine and mupirocin (Patel JB, 2009).

- Given the limited evidence, careful consideration should be given to risk, benefits and outcomes to individuals and wider community (family, ward, institution etc) before offering decolonisation to contacts.
- The decision to decolonise contacts with or without screening in the case of a PVL-S. *aureus* outbreak within a healthcare setting will be risk assessed by the local Incident Management Team (IMT).
- If the decision made concludes that decolonisation with or without screening is required, the local policy for MRSA decolonisation may be used
- If the decision is made to offer decolonisation to close contacts of index cases who may have had PVL infections, it is prudent to wait until infection has resolved before starting and then start all contacts simultaneously.
- Decolonisation of neonates and children, especially premature neonates, is more difficult and unstandardised. When required, nasal mupirocin may be used as may aqueous based antiseptic solutions, such as chlorhexidine. Octenisan has been used as an alternative to chlorhexidine, but this is now has a cosmetic licence and requires an individual risk assessment as the company cannot recommend its use in children under 3 years of age. This topical agent requires a contact time of at least one minute and may cause a temperature drop in neonates. Further advice will be available from local infection prevention and control staff.
- Patients should be given information leaflets for PVL (<u>Appendix C on page 32</u>) describing how to minimise cross-infection and general information about PVL. Decolonisation protocols are not included in this document as the local MRSA decolonisation protocols and patient information can be used. However, some supplementary information is available in <u>Appendix B on page 31</u>.
- In patients with dermatological conditions it is important to seek dermatological opinions, if there are ongoing issues with skin integrity (Fogo, et al 2011).
- Advice from Human Resources and Occupational Health must be sought when dealing with screening or exclusion of staff from work, following current Scottish guidance. 'NHS HDL (2006)31 Healthcare Associated Infection (HAI): Human Resources Policy for Staff Screening during Incidents and Outbreaks' available at http://www.sehd.scot.nhs.uk/mels/HDL2006_31.pdf. The decision to screen and/or decolonise staff during a cluster or outbreak will be made by the local IMT.
- The need for rescreening post decolonisation is unclear, but local MRSA screening protocols can be used. It may be appropriate for those patients who are vulnerable to infection (e.g. dialysis patients) or those who pose a risk of onward transmission (healthcare workers, those in closed communities).

7. Clusters and Outbreaks of PVL-S. aureus Infections

Health Protection Teams (HPTs) should be informed of possible clusters or outbreaks of confirmed PVL-S. *aureus* infection if identified by clinical, laboratory or other healthcare professionals.

 HPTs will review the circumstances of each cluster/outbreak, and, as a multi-disciplinary team, will decide on the most appropriate response, using formats such as a Problem Assessment Group (PAG), with representatives including staff from the affected area, Microbiologists etc. and others as required. Please refer to the Scottish Government/HPN (2011) 'Management of Public Health Incidents: Guidance on the Roles and Responsibilities of NHS led Incident Management Teams' available at http://www.scotland.gov.uk/ Publications/2013/08/6455/downloads.

Clusters and outbreaks can occur in:

- those involved in gyms or 'close contact' sports, such as wrestling and rugby (see section 3.2 on page 8);
- a 'closed' community where there may be potential for onward transmission (e.g. prison, military camp, carehome, healthcare setting); as well as
- households.
- nursery schools, these will have to be managed in consultation with Health Protection Teams on a case by case basis.

However, paucity of epidemiological data means that it is not always possible to identify a community-based cluster or outbreak. Some HPTs may use postcode data to try and track potential household outbreaks.

In a healthcare setting such as a hospital, it may be easier to identify when there are more than two cases associated by place or time. In this situation, the local Infection Prevention and Control Team will undertake review and response (see section 8 on page 20).

- The National Infection Prevention and Control Manual provides guidance to all those involved in care provision and should be adopted for infection prevention and control practices and procedures.
- Standard Infection Control Precautions must continue to be applied and additional TBPs considered and implemented as set out in the National Infection Prevention and Control Manual: <u>http://www.hps.scot.nhs.uk/haiic/</u> ic/nationalinfectionpreventionandcontrolmanual.aspx.

Further advice:

- RCPI Clinical Advisory Group on HCAI & AMR, Subgroup MRSA Guidelines
- Committee (April 2013). Eradication of MRSA carriage (decolonisation). In: The Control and Prevention of MRSA. [Online] Available from: http://www.hpsc.ie/A-Z/MicrobiologyAntimicrobialResistance/ EuropeanAntimicrobialResistanceSurveillanceSystemEARSS/ ReferenceandEducationalResourceMaterial/SaureusMRSA/Guidance/ File,14478,en.pdf [Accessed 13 August 2013].
- Useful advice can be found in the recommendations given at the 'Guidelines for the control and prevention of Meticillin-resistant *Staphylococcus aureus* (MRSA) in healthcare facilities' (Coia, 2006).

For household outbreaks of infection, patients will be treated by their own GP. However, there may be situations where different GPs are caring for different members of the household and in this situation co-ordination of treatment and, if required, decolonisation should be attempted; as would be the case for households undergoing treatment for scabies etc.

Recommendation (for Research):

 Consider enhanced surveillance programme for laboratory confirmed cases of PVL with a view to enhancing existing knowledge on epidemiology of this in Scotland.

8. Infection Prevention and Control (IPC) Measures

8.1 Infection Prevention and Control Measures for Hospitalised Patients

Hospitals have policies and procedures which deal with MRSA and these are generally appropriate for the control of PVL-S. *aureus* cases and outbreaks.

 Standard Infection Control Precautions must continue to be applied and additional TBPs considered and implemented as set out in the National Infection Prevention and Control Manual: <u>http://www.hps.scot.nhs.uk/haiic/</u> ic/nationalinfectionpreventionandcontrolmanual.aspx.

During an outbreak investigation, the following tools are available:

- The 'Hospital Infection Incident Assessment (HIIA) Tool' (also referred to as HIIAT). Available online: <u>http://www.documents.hps.scot.nhs.uk/hai/infection-control/toolkits/hiiat-2011-10.pdf</u>;
- 'Healthcare Outbreak Algorithm For Patient, Healthcare Worker and Visitor (PHV) Safety to optimise patient, healthcare worker and visitor safety during outbreak investigations'. Available online: <u>http://www.documents.hps.scot.</u> <u>nhs.uk/hai/infection-control/toolkits/hospital-outbreak-management-2013-05.</u> <u>pdf</u>.

The Scottish MRSA Reference Laboratory can be contacted for expert advice (contact details at <u>http://www.smrsarl.scot.nhs.uk/</u>).

8.1.1 Infection Prevention and Control (IPC) Measures in Relation to Patients with Necrotising Pneumonia

Standard Infection Control Precautions (SICPs) must continue to be applied. In addition: HCWs should wear facial and respiratory protective equipment e.g. a fluid repellent surgical face mask with integral eye protection if there is likely splashing or spraying of blood/body fluids from patient contact or procedure e.g. during intubation and any respiratory care of a patient with known/suspected necrotising pneumonia (refer to the National Infection Prevention and Control Manual: <u>http://www.hps.scot.nhs.uk/haiic/ic/</u> <u>nationalinfectionpreventionandcontrolmanual.aspx</u>.

If a HCW has direct contact with respiratory secretions from a PVL-positive patient they should be advised to seek advice from the local Occupational Health Service. If a HCW subsequently becomes unwell (fever, flu-like illness, sore throat) 3 to 7 days post exposure they should be advised to seek medical attention.

8.1.2 IPC Measures in Relation to Hospital-acquired Infections

If a case of PVL-S. aureus infection was acquired or there is the possibly it was acquired, in hospital, a risk assessment needs to be undertaken by the Infection Prevention & Control Team and infection prevention and control measures put in place (as above).

If there are linked cases suggesting an outbreak or cross infection, screening and decolonisation of other patients and staff (if the epidemiology suggests this) should be considered by the Incident Management Team (IMT).

The Microbiology department should search its database for *S. aureus* infections with an alert antibiogram that may be related, and any isolates, if still available, sent to the SMRSARL for PVL-testing. This will help to ascertain any unidentified clusters of cases in the hospital.

It is good practice that any significant learning points as a consequence of postincident or outbreak review are shared locally and nationally.

8.1.3 Occupational Health

A HCW with a proven PVL-S. *aureus* infection should not work until the acute infection has been resolved and until at least 48 hours of a five day decolonisation regimen has been completed. A risk assessment will need to be undertaken with input from Infection Prevention and Control staff or Microbiologist depending on the nature of the work. Enquiries regarding PVL-S. *aureus*-related disease in close household/sexual contacts of the staff member should be made, so decolonisation and treatment can be offered simultaneously, if required.

Follow up samples, following topical decolonisation, are advised as for MRSA guidelines (three screens, one week apart). Unlike MRSA, staff who are found to have PVL-*S*. *aureus* are likely to have acquired the infection in the community, and hence re-colonisation may occur from a close contact. Therefore, even if screens have been negative, staff should stop working and seek both treatment and Occupational Health advice if a further skin lesion develops.

If, despite two courses of decolonisation treatment, a staff member remains colonised, they should be able to continue work providing they are not implicated in hospital transmission of PVL-S. *aureus* infection and they cease working as soon as an infected skin lesion develops. This will require individual assessment of risks to patients and the staff member and will require multidisciplinary input including Occupational Health, Infection Prevention and Control staff and the line manager.

Occupational health issues will be the responsibility of the patient's employer in conjunction with the GP treating the infection.

8.2 Infection Prevention and Control Measures for Household Settings

The key principles of preventing and controlling the spread of infection in the household setting centre on:

- early suspicion of infection, with rapid diagnosis and appropriate treatment;
- the natural course of the infection with relapses;
- environmental survival of the bacteria.

Measures that can be put in place to reduce spread within a household:

- ensuring lesions are covered with clean, dry dressings, which are changed and disposed of as soon as discharge seeps to the surface;
- personal hygiene and good skin care (particularly those with eczema);
- using separate towels and not sharing personal items such as razors, toothbrushes, face cloths etc.; replacement of personal utensils e.g. toothbrushes, combs;
- ensuring laundry of towels, bedlinen, clothing etc using a hot wash (60°C), where possible - daily;
- regular household cleaning daily where possible;
- avoiding communal and recreational settings until lesions are healed if they cannot be adequately contained by a dressing; certain activities such as use of gym, sauna, swimming pool use, massages, manicures or similar, should be avoided until the lesions have healed.

Further advice:

- those who work in occupations where they might pose a risk of infection to others, such as healthcare workers; carers in nurseries, residential or care homes or similar; or food handlers, should not attend work until the lesions have healed;
- those who have eczema or a more generalised skin condition should remain off work or school until treatment has been completed;
- children can go to school if they can understand the importance of good hand hygiene, and can keep their infected skin covered with a clean dry dressing which will stay dry and in place until the end of the school day.
- children attending nurseries will require individual assessment in terms of suitability to return to nursery.

References

AGREE Collaboration (2009). Appraisal of Guidelines for Research and Evaluation. (AGREE) Instrument II. <u>http://www.agreetrust.org/resource-centre/training/</u>

Al-Talib H, CY Yean, et al. (2009). A pentaplex PCR assay for the rapid detection of methicillin-resistant *Staphylococcus aureus* and Panton-Valentine Leucocidin. *BMC Microbiol* 9: 113.

Ammerlaan HS, Kluytmans JA, Wertheim HF, Nouwen JL, Bonten MJ. Eradication of methicillinresistant *Staphylococcus aureus* carriage: a systematic review. *Clin Infect Dis* 2009 Apr 1;48(7):922-30.

Badiou C, O Dumitrescu, et al. (2010). Rapid detection of Staphylococcus aureus Panton-Valentine leukocidin in clinical specimens by enzyme-linked immunosorbent assay and immunochromatographic tests. Journal of Clinical Microbiology 48(4):1384-1390.

Bae IG, GT Tonthat, et al. (2009). Presence of genes encoding the pantonvalentine leukocidin exotoxin is not the primary determinant of outcome in patients with complicated skin and skin structure infections due to methicillinresistant *Staphylococcus aureus*: results of a multinational trial. *Journal of Clinical Microbiology* 47(12): 3952-3957.

Baggett HC, TW Hennessy, et al. (2004). Community-onset methicillin-resistant Staphylococcus aureus associated with antibiotic use and the cytotoxin Panton-Valentine leukocidin during a furunculosis outbreak in rural Alaska. Journal of Infectious Diseases 189(9): 1565-1573.

Bittar F, Z Ouchenane, et al. (2009). MALDI-TOF-MS for rapid detection of staphylococcal Panton-Valentine leukocidin. International Journal of Antimicrobial Agents 34(5):467-470.

Blaine KP, MJ Tuohy, et al. (2010). Progression to bacteremia in critical care patients colonised with methicillin-resistant *Staphylococcus aureus* expressing Panton-Valentine leukocidin. *Diagnostic Microbiology & Infectious Disease 68*(1): 28-33.

Bocchini CE, KG Hulten, et al. (2006). Panton-Valentine leukocidin genes are associated with enhanced inflammatory response and local disease in acute hematogenous *Staphylococcus aureus* osteomyelitis in children. *Pediatrics* 117(2): 433-440.

Carre N, F Sillam, et al. (2008). Staphylococcus aureus carrying Panton-Valentine leukocidin genes nasal colonisation and skin infection: screening in case of outbreak in a school environment. Medecine et Maladies Infectieuses 38(9): 483-488.

Chini V, E Petinaki, et al. (2006). Spread of Staphylococcus aureus clinical isolates carrying Panton-Valentine leukocidin genes during a 3-year period in Greece. Clinical Microbiology & Infection 12(1): 29-34.

Chua K, Laurent F, et al. (2011). Not Community-Associated Methicillin_resistant Staphylococcus aureus (CA-MRSA) A clinician's guide to community MRSA – its evolving antimicrobial resistance and implications for therapy. *Clinical Infectious Diseases* 52 (1): 99-101. Coia JE, Duckworth GJ, Edwards DI, et al (2006). Guidelines for the control and prevention of meticillin-resistant *Staphylococcus aureus* (MRSA) in healthcare facilities. J Hosp Infect. 2006 May 63 Suppl 1:S1-44.

Cunnington A, T Brick, et al. (2009). Severe invasive Panton-Valentine Leucocidin positive Staphylococcus aureus infections in children in London, UK. J Infect. 59(1): 28-36.

Cruciani M, Gatti G, Lazarini L. (1996) Penetration of vancomycin into human lung tissue. Antimicrob Ag Chemother 38: 865-9.

Dailiana ZH, N Rigopoulos, et al. (2008). Clinical and epidemiological features of upperextremity infections caused by *Staphylococcus aureus* carrying the PVL gene: a four-year study in Greece. *Medical Science Monitor* 14(10): CR511-514.

De Angelis G, Laurent F et al. (2011) Treatment of Skin and Soft Tissue Infections Due to Community-Associated Methicillin-Resistant Staphylococcus aureus in Europe: The Role of Trimethoprim-sulfamethoxazole Clin Infect Dis. 52(12): 1471-1472

Dellinger RP, Levy MM, Carlet JM, Bion J, Parker MM, Jaeschke R, Reinhart K, Angus DC, Brun-Buisson C, Beale R, Calandra T, Dhainaut JF, Gerlach H, Harvey M, Marini JJ, Marshall J, Ranieri M, Ramsay G, Sevransky J, Thompson BT, Townsend S, Vender JS, Zimmerman JL, Vincent JL. (2008). International guidelines for management of severe sepsis and septic shock. *Crit Care Med* 36:296-327.

Deurenberg RH, C Vink, et al. (2004). Rapid detection of Panton-Valentine leukocidin from clinical isolates of *Staphylococcus aureus* strains by real-time PCR. *FEMS Microbiology Letters* 240(2): 225-228.

Dohin B, Y Gillet, et al. (2007). Pediatric bone and joint infections caused by Panton-Valentine leukocidin-positive *Staphylococcus aureus*. Pediatric Infectious Disease Journal 26(11): 1042-1048.

Ellington MJ, Perry C, Ganner M, Warner M, McCormick Smith I, Hill RL, et al. Community Associated MRSA with the PVL toxin in England and Wales: Eur J Clin Microbiol. Infect Dis. 2009 Sep;28(9):1113-21.

Enayet I, Nazeri A, Johnson LB, Riederer K, Pawlak J, Saravolatz LD. (2006) Community-associated methicillin-resistant *Staphylococcus aureus* causing chronic pneumonia. *Clin Infect Dis* 42: 357-9.

Etienne J, Dumitrescu O (2009). Panton-Valentine leucocidin associated *Staphylococcus aureus* infections. *BMJ* 339: b4083.

Fogo A, Kemp N, Morris-Jones R. (2011) BMJ 2011;343:d5343.

Gauduchon V, G Cozon, et al. (2004). Neutralization of Staphylococcus aureus Panton Valentine leukocidin by intravenous immunoglobulin in vitro. Journal of Infectious Diseases 189(2): 346-353.

Gbaguidi-Haore, HM Thouverez, et al. (2009). Usefulness of antimicrobial resistance pattern for detecting PVL- or TSST-1-producing meticillin-resistant *Staphylococcus aureus* in a French university hospital. *Journal of Medical Microbiology* 58 (Pt 10): 1337-1342.

Gillet, YP Vanhems, et al. (2007). Factors predicting mortality in necrotizing community acquired pneumonia caused by *Staphylococcus aureus* containing Panton-Valentine leukocidin. *Clinical Infectious Diseases* 45(3): 315-321.

GRADE working group, T. (2004). Grading quality of evidence and strength of recommendations (abridged version). *British Medical Journal* 328: 1490-1494.

Hawkins G, Stewart S, Blatchford O, Reilly J. (2011) Should healthcare workers be screened routinely for meticillin-resistant *Staphylococcus aureus*? A review of the evidence. *J Hosp Infect*. 77(4):285-9

Health Protection Agency (HPA), U. L. A. R. S. (2010). Management of PVL-Staphylococcus aureus - Recommendations for practice.

Health Protection Agency (HPA), U. (2008). Guidance on the Diagnosis and Management of PVL-associated *Staphylococcus aureus* infections (PVL-S. *aureus*) in England. Second Edition: 7 November 2008. First Edition published: 15 August 2008.

Holmes A, M Ganner, et al. (2005). Staphylococcus aureus isolates carrying Panton Valentine leucocidin genes in England and Wales: frequency, characterization, and association with clinical disease. Journal of Clinical Microbiology 43(5): 2384-2390.

Jahamy H, R Ganga, et al. (2008). Staphylococcus aureus skin/soft-tissue infections: the impact of SCCmec type and Panton-Valentine leukocidin. Scandinavian Journal of Infectious Diseases 40(8): 601-606.

Karahan ZC, A Tekeli, et al. (2008). Investigation of Panton-Valentine leukocidin genes and SCCmec types in clinical Staphylococcus aureus isolates from Turkey. *Microbial Drug Resistance-Mechanisms Epidemiology & Disease* 14(3): 203-210.

Köck R, Becker K, Cookson B, van Gemert-Pijnen JE, Harbarth S, Kluytmans J, Mielke M, Peters G, Skov RL, Struelens MJ, Tacconelli E, Navarro Torné A, Witte W, Friedrich AW. (2010) Methicillin-resistant *Staphylococcus aureus* (MRSA): burden of disease and control challenges in Europe. *Euro Surveill*. 2010;15(41):pii=19688. <u>http://www.eurosurveillance.org/ViewArticle.aspx?ArticleId=19688</u>

Lamer C, de Beco V, Soler P. (2002) Analysis of vancomycin entry into pulmonary lining fluid by bronchoalveolar lavage in critically ill patients. *Antimicrob Ag Chemother* 46:1475-80.

Liassine N, R Auckenthaler, et al. (2004). Community-acquired methicillin-resistant Staphylococcus aureus isolated in Switzerland contains the Panton-Valentine leukocidin or exfoliative toxin genes. Journal of Clinical Microbiology 42(2):825-828.

Libert N, E Batjom, et al. (2009). Antitoxin treatments for necrotizing pneumonia due to Panton-Valentine leukocidin-secreting *Staphylococcus aureus*. Medecine et Maladies Infectieuses 39(1): 14-20.

Linde H, N Lehn (2005). Infections with methicillin-resistant Staphylococcus aureus: impact of Panton-Valentine leukocidin. Deutsche Medizinische Wochenschrift 130(42): 2397-2401.

Liu C, Bayer A, et al. (2011). Clinical Practice guidelines by the Infectious Disease Society of America for the treatment of Methicillin-resistant *Staphylococcus aureus* infections in adults and children.*Clin Infect Dis* 52 (3); e18-55.

Lo WT, CS Tang, et al. (2009). Panton-Valentine leukocidin is associated with exacerbated skin manifestations and inflammatory response in children with community-associated staphylococcal scarlet fever. *Clinical Infectious Diseases* 49(7): e69-75.

Ma X, T Ito, et al. (2008). Two different Panton-Valentine leukocidin phage lineages predominate in Japan. Journal of Clinical Microbiology 46(10): 3246-3258.

Martinez-Aguilar G, A Avalos-Mishaan, et al. (2004). Community-acquired, methicillinresistant and methicillin-susceptible *Staphylococcus aureus* musculoskeletal infections in children. *Pediatric Infectious Disease Journal* 23(8): 701-706.

Masiuk H, K Kopron, et al. (2010). Association of recurrent furunculosis with Panton-Valentine leukocidin and the genetic background of *Staphylococcus aureus*. Journal of Clinical Microbiology 48(5): 1527-1535.

McGrath B, Rutledge F, Broadfiel E. (2008) Necrotising pneumonia, Staphylococcus aureus and Panton-Valentine leukocidin. *Journal of the Intensive Care Society* 2008; 9 (2): 170-172.

Morgan MS. (2007). Diagnosis and treatment of Panton-Valentine leukocidin (PVL) associated staphylococcal pneumonia. International Journal of Antimicrobial Agents 30(4): 289-296.

Muttaiyah S, G Coombs, et al. (2010). Incidence, risk factors, and outcomes of Panton-Valentine leukocidin-positive methicillin-susceptible *Staphylococcus aureus* infections in Auckland, New Zealand. *Journal of Clinical Microbiology* 48(10):3470-3474.

Nakagawa S, I Taneike, et al. (2005). Gene sequences and specific detection for Panton-Valentine leukocidin. *Biochemical & Biophysical Research Communications* 328(4):995-1002.

Nathwani D, Morgan M, Masterton RG, Dryden M, Cookson BD, French G, Lewis D on behalf of the British Society for Antimicrobial Chemotherapy Working Party on community-onset MRSA Infections. (2008). Guidelines for UK practice for the diagnosis and management of methicillin-resistant *Staphylococcus aureus* (MRSA) infections presenting in the community. J Antimicrob Chemother 61(5): 976-994 <u>http://jac.oxfordjournals.org/content/61/5/976.full</u>

Nhan TX, Bes M, Meugnier H, Toko L, Julienne G, Thiolet JM, Tillier C, Tessier S, Baverel J, Conscience B, Lavigne JP, Laurent F, Etienne J, Vandenesch F, Tristan A. (2012) ST93-Queensland community-acquired meticillin-resistant *Staphylococcus aureus* clone in France: outbreak in a scout camp and sporadic cases, July to August 2012. *Euro Surveill*. 2012; 17(44):pii=20307. <u>http://www.eurosurveillance.org/</u> <u>ViewArticle.aspx?ArticleId=20307</u>

Oishi K, T Baba, et al. (2008). A latex agglutination assay for specific detection of Panton–Valentine leukocidin. Journal of Microbiological Methods 75: 411-415.

Orendi JM, N Coetzee, et al. (2010). Community and nosocomial transmission of Panton-Valentine leucocidin-positive community-associated meticillin-resistant *Staphylococcus aureus*: implications for healthcare. Journal of Hospital Infection 75(4): 258-264.

Panton PN, Valentine FCO. Staphylococcal toxin. Lancet. 1932: 219 :506-508.

Papenburg J, P Fontela, et al. (2009). Panton-Valentine leukocidin in pediatric community-acquired Staphylococcus aureus infections. Clinical & Investigative Medicine - Medecine Clinique et Experimentale 32(5): E352-359.

Patel JB, Gorwitz RJ, Jernigan JA. (2009) Mupirocin resistance. Clin Infect Dis 49:935–941.

Patel M, Waites KB, Hoesley CJ, Stamm AM, Canupp KC, Moser SA. (2008). Emergence of USA300 MRSA in a tertiary medical centre: implications for epidemiological studies. J Hosp Infect 68: 208-213.

Rajendran PM, D Young, et al. (2007). Randomized, double-blind, placebocontrolled trial of cephalexin for treatment of uncomplicated skin abscesses in a population at risk for community-acquired methicillin-resistant *Staphylococcus aureus* infection. *Antimicrobial Agents & Chemotherapy* 51(11): 4044-4048.

Reilly JS, Stewart S, Christie P, Allardice G, Smith A, Masterton R, Gould IM, Williams C. Scottish Government Health Directorate (Funder) (2009) Universal screening for meticillin-resistant *Staphylococcus aureus*: interim results from the NHS Scotland pathfinder project. *Journal of Hospital Infection*, 74 (1). pp. 35-41. ISSN 0195-6701. http://strathprints.strath.ac.uk/15076/1/reilly_jhi_2010_74_35-41(1).pdf

Reischl U, MJ Tuohy, et al. (2007). Rapid detection of Panton-Valentine leukocidinpositive Staphylococcus aureus by real-time PCR targeting the lukS-PV gene. European Journal of Clinical Microbiology & Infectious Diseases 26(2): 131-135.

Renwick L, A Hardie, et al. (2008). Detection of meticillin-resistant Staphylococcus aureus and Panton-Valentine leukocidin directly from clinical samples and the development of a multiplex assay using real-time polymerase chain reaction. European Journal of Clinical Microbiology & Infectious Diseases 27(9): 791-796.

Sdougkos GV, Chini, et al. (2007). Methicillin-resistant Staphylococcus aureus producing Panton-Valentine leukocidin as a cause of acute osteomyelitis in children. Clinical Microbiology & Infection 13(6): 651-654.

Shallcross LJ, K Williams, et al. (2010). Panton-Valentine leukocidin associated staphylococcal disease: a cross-sectional study at a London hospital, England. *Clinical Microbiology & Infection* 16(11): 1644-1648.

Shallcross LJ, Fragaszy E, Johnson AM, Hayward AC. (2013). The role of the Panton-Valentine leucocidin toxin in staphylococcal disease: a systematic review and meta-analysis. *Lancet Infect Dis* 13: 43-54.

Shallcross LJ, Mbeledogu CN, Hayward AC. Should we screen and decolonise contacts of patients with Panton Valentine leukocidin associated *Staphylococcus aureus* infection? *BMJ* 2011: 343: d5479.

Srinivasan A, S Seifried, et al. (2009). Panton-Valentine leukocidin-positive methicillinresistant *Staphylococcus aureus* infections in children with cancer. *Pediatric Blood & Cancer 53*(7): 1216-1220.

Stevens DL, Ma Y, McIndoo E, Wallace RJ, Bryant A. (2007) Impact of antibiotics on expression of virulence-associated exotoxin genes in methicillin-sensitive and methicillin-resistant *Staphylococcus aureus*. J Infect Dis. 195: 202-11.

Tang YW, A Kilic, et al. (2007). StaphPlex system for rapid and simultaneous identification of antibiotic resistance determinants and Panton-Valentine leukocidin detection of staphylococci from positive blood cultures. Journal of Clinical Microbiology 45(6): 1867-1873.

Thuong TC, Tho ND, et al. (2007). An Outbreak of Severe Infections with Community-Acquired MRSA carrying the Panton-Valentine Leukocidin following Vaccination. *PLosOne* September 2007 (9 e822): 1-6.

Vardakas KZ, DK Matthaiou, et al. (2009). Comparison of community-acquired pneumonia due to methicillin-resistant and methicillin-susceptible Staphylococcus aureus producing the Panton-Valentine leukocidin. International Journal of Tuberculosis & Lung Disease 13(12): 1476-1485.

Vourli S, H Vagiakou, et al. (2009). High rates of community-acquired, Panton-Valentine leukocidin (PVL)- positive methicillin-resistant S. aureus (MRSA) infections in adult outpatients in Greece. Bulletin Europeen sur les Maladies Transmissibles European Communicable Disease Bulletin 14(2).

Wagenlehner FME, KG Naber, et al. (2007). Management of a large healthcare associated outbreak of Panton-Valentine leucocidin-positive meticillin-resistant Staphylococcus aureus in Germany. Journal of Hospital Infection 67(2): 114-120.

Zamora J, Abraira V, et al. (2006). Meta-Disc: a software for meta-analysis of test accuracy data. BMC Medical Research Methodology (6): 31.

Appendix A: Quick Reference Guide for PVL-S. aureus for Primary Care (Adapted from HPA Chart 18/5/09)

Quick Reference Guide for PVL-S. aureus for Primary Care

- Usually skin infections, but also severe invasive infection in otherwise healthy young individuals.
- Infections can be recurrent over several months, affect several household members and disrupt work, school and social life.

Characteristics of infection with PVL-S. aureus

Recurrent skin infections

- Boils, carbuncles, cellulitis.
- Pain and erythema out of proportion to severity of signs.
- Necrosis may be present.

Invasive infections

- Necrotising pneumonia after a flu-like illness.
- Necrotising fasciitis.
- Purpura fulminans.
- Osteomyelitis, septic arthritis, pyomyositis.

Risk factors and groups at high risk for PVL-S. aureus

- Closed communities with close contact e.g. care homes, healthcare premises, prisons etc.
- Contact sports (rugby, wrestling etc.).
- Military training camps.
- Gyms.
- Travel outwith UK.
- Contact with a known case.

Factors contributing to cross infection:

- Crowding.
- Close contact (household/sexual).
- Cuts/compromised skin integrity.
- Contamination of environment.

When should I investigate for PVL-S. aureus? NB: State PVL on request form

- Recurrent boils and abscesses.
- Necrotising skin and soft tissue infection.
- More than one typical case in household or closed community.
- Community acquired necrotising/haemorrhagic pneumonia (sputum, swabs, and refer to hospital immediately).

What bacteriology samples should I take? Make sure PVL I son request

- Swabs submitted in locally specified manner of skin lesions/breaks, pus etc.
- Others depending on nature of infection.

When and how do I treat PVL-infections?

- Abscesses should be incised and drained. Send pus for culture if PVL suspected.
- Cellulitis, large abscesses pending drainage give flucloxacillin orally 5-7 days.
- Clindamycin or cotrimoxazole can be used for penicillin allergic patients.

If MRSA is suspected or confirmed empiric MRSA treatment as per your local antibiotic protocols.

If there is severe SSTI with systemic symptoms or pneumonia refer to hospital immediately.

When should I offer decolonisation? (Limited evidence, expert consensus)

- To index cases after lesions have healed (no re-screening unless high risk group).
- To household contacts of necrotising/haemorrhagic pneumonia (no screening needed).
- If index case is a special risk to others (healthcare worker) with re-screening.
- Give patient local information leaflet on decolonisation protocol.
- Only after infection is resolved.

Follow up

- Should not attend work/school or nursery whilst lesion is active or cannot be covered.
- Healthcare staff should contact their Occupational Health Department for assessment.
- Patient information leaflet should be provided for those with confirmed PVL.

Appendix B: Patient information – Decolonisation General Information

Decolonisation procedure for PVL-Staphylococcus aureus (PVL-S. aureus)

The purpose of decolonisation is to try to reduce the number of bacteria in carriage sites; moist areas of the body, such as nose, armpits and groin, to reduce relapses of infection, spread to close contacts or risk of a wound infection before an operation for example. Generally you will be advised to have any infection treated and settled before decolonisation starts. Sometimes if a close contact has a **very** serious infection you may be advised to have the decolonisation earlier.

The decolonisation consists of two parts (1) an antiseptic skin wash instead of normal soap and (2) an antiseptic nose cream for 5 days. You will be given details on how to use this.

This will not get rid of the bacteria if they are in other carriage sites such as the gut or throat and it may not be completely successful in all cases. However, most people with PVL-S. *aureus* carry it in the nose and sometimes reducing the number of the germs can help to break an ongoing cycle of infection between close contacts.

It is not of proven value in PVL management, but similar approaches have been used for many years to reduce MRSA colonisation and generally it is safe and well tolerated.

This treatment will only be started once any infection is treated for the best chance of success.

Reducing spread of the bacteria within the households during the antiseptic treatment:

- Clean your hands often (before touching young children, preparing food, eating, doing housework and after going to the toilet). Keep fingernails short.
- If possible change underwear, bedding, clothing and towels daily. If you can't manage this try at least to do it on days 2 and the last day the treatment.
- Use fresh wash cloth daily or use a disposable one and throw it away every day; or avoid using them at all. Don't share toothbrushes, hair brushes, towels, washcloths etc.
- Pay particular attention to armpits, groin, 'down below' when using the antiseptic soap. If possible use the antiseptic soap in your hairline/hair (avoid if hair is coloured or permed).
- If your skin gets itchy with the antiseptic body wash stop using it and contact your GP.
- If your household contacts are also to be given the antiseptic treatment it's useful if you all start it together.

Appendix C: Patient Information PVL-Staphylococcus aureus

What is PVL-Staphylococcus aureus (PVL-S. aureus)?

Staphylococcus aureus is a germ that commonly lives on healthy skin. About one third of healthy people carry it quite harmlessly, usually on carriage sites; moist areas of the body, such as nose, armpits and groin, this is known as colonisation. A small number of S. aureus can produce a toxin called Panton-Valentine Leucocidin (PVL) and they are known as PVL-S. aureus.

Less than 2% of S. aureus produce this toxin and it can be found in MRSA or meticillin sensitive *Staphylococcus aureus* (MSSA). PVLs are mainly found in the community rather than in hospitals. Both MSSA and MRSA types are sensitive to a variety of antibiotics – many of which are available as tablets.

What type of illness does it cause?

PVL strains may cause no infection (carriers) or they may cause infections like boils or abscesses – these may well occur several times. Very rarely they can cause more serious infection of the lungs or bones which may require hospital treatment.

How do you catch PVL-S. aureus?

Anyone can get a PVL-S. *aureus* infection. Infection can also occur in fit, healthy people during skin-to-skin contact with someone who has the germ, for example within a family, during contact sports like rugby, or from contaminated surfaces for example shared gym equipment, razors, towels etc. Damaged skin can be more prone to infection with any S. *aureus*, including PVL.

How is PVL-S. aureus treated?

Boils and abscesses should be drained of pus where possible. Some infections (MSSA or MRSA) may be treated with a course of antibiotic tablets and there are several different choices available.

In certain cases the doctor may suggest the use of antiseptic washes and nose ointments to reduce the number of germs present on the skin after infection has healed. This may help reduce the chance of repeated infection or break a cycle of infection occurring in different household members. It may not get rid of the germ completely. In cases where there are other household or close contacts suffering infections, the antiseptic wash and nose ointment may be suggested for everyone.

How do I prevent passing PVL-S. aureus to other people?

Keep the infected areas covered with clean, dry dressings or plasters. Change these regularly or as soon as you see seepage to the surface of the dressing/ plaster. Wash your hands before and after changing the dressings.

Do not touch, poke or squeeze infected skin. This transfers the germs to your hands and can push them deeper into the skin.

Cover your nose and mouth with a tissue when you cough or sneeze, particularly if you have a cold, because the germs can live in your nose. Throw the tissue in the bin at once and then wash your hands.

Try and keep personal items like towels, razors, toothbrushes, etc for your own use. Wash towels frequently at the highest temperate the materials will allow.

Can I go to work or school when I have a PVL-S. aureus infection?

- You should not work as a carer in a nursery, hospital, residential or care home or similar place until your skin has healed and you have permission to return to work from your local Occupational Health Department, GP or manager.
- You should not work in the food industry, e.g. waitress, chef, food production, until your skin has healed and you have permission to return to work from your local Occupational Health Department, GP or manager.
- You may carry on with other types of work, provided you keep infected skin areas covered with clean, dry dressings. If you are not sure about working, contact your local Occupational Health Department or GP.
- Children can go to school if they are old enough to understand the importance of good hand hygiene, and if their infected skin is covered with a clean dry dressing which will stay dry and in place until the end of the school day. Children should not take part in contact sports, or use communal gym equipment until their skin is healed. The GP's advice is essential and school management should be informed.
- People with eczema or a more generalised skin condition should take advice from their GP whether to remain off work or school. You need to continue treating your skin to keep it in good condition. In the long term this helps to reduce the risk of spread of PVL-S. *aureus*.

Can I go to swimming pools, gyms or sports facilities when I have a PVL-S. aureus infection?

You should not use communal facilities for example gym equipment, saunas, swimming pools, or have a massage, manicure or similar until your skin has healed.

How do I prevent becoming infected again?

If you are found to carry PVL-S. aureus persistently on your skin or nose, or if you suffer from repeated infections, you may be given a course of skin and nose disinfectant treatment. Sometimes the skim treatment will be extended to your household or close contacts. In these circumstances it is important that all affected people in a household or social group are treated at the same time.

If you have a further infection of any type, if you are admitted to hospital unexpectedly, or if you are going to be admitted to hospital for an operation, always tell the doctor or nurse looking after you that you have had a PVL-S. *aureus* infection. This will ensure that you receive appropriate treatment.

Appendix D: Guideline Development Group (GDG) Membership

Gabby Phillips – NHS Tayside Consultant Microbiologist (GDG Chair)

Oliver Blatchford – Health Protection Scotland Consultant Public Health Medicine

Barry Cookson – Health Protection Agency Director, Laboratory of Healthcare Associated Infection

Pamela Davidson – NHS Tayside Senior Infection Control Nurse

Martin Donaghy – Health Protection Scotland Divisional Medical Director

Giles Edwards – NHS Greater Glasgow & Clyde Consultant Microbiologist

Carol Fraser – Scottish Government Health Policy Unit

Pamela Joannidis – NHS Greater Glasgow & Clyde Lead Nurse Consultant

Laura Jones – NHS Lothian Consultant Pediatrician

Ann Mathieson – NHS Greater Glasgow & Clyde Health Protection Nurse Specialist

Vincent McKeown – NHS Greater Glasgow & Clyde Health Protection Nurse Specialist

Dilip Nathwani – NHS Tayside Consultant Physician

Ida Prantner – Health Protection Scotland EPIET Fellow

Mary Quinn – NHS Fife Health Protection Nurse Specialist & HPN Steering Group Chair

Janette Richards – NHS Lothian Infection Control Nurse

Matthew Riddell – Health Protection Agency Programme Manager

Lisa Ritchie – Health Protection Scotland Nurse Consultant Infection Control

Alex Sanchez-Vivar – Health Protection Scotland

EPIET Fellow

Charles Saunders – NHS Fife

Consultant Public Health Medicine

Jacqueline Sneddon – Healthcare Improvement Scotland Project Lead - Scottish Antimicrobial Prescribing Group

Liz Stokle – Health Protection Agency HCAI & AMRS Programme Lead/Nurse Epidemiologist

Julie Wilson – Health Protection Scotland Epidemiologist

Appendix E: HPN Guideline Feedback form

| | Section A – About the Document (Guideline) |
|----|---|
| | Guideline Title: |
| | Author: |
| | Publisher: |
| | Date of Publication: |
| | Section B – About the Evaluation |
| | Reviewer's Name: |
| | Reviewer's Occupation: |
| | Reviewer's Organisation: |
| | Reviewer's Contact Email Address: (Optional) |
| | Date of Evaluation: |
| | Section C – Comments |
| • | Does the Guideline meet your needs/inquiry at the time of evaluation? (Please explain why this is the case.) |
| 2. | Is there anything lacking in the Guideline? (Please explain.) |
| - | Do you have any other comments? |